

A Novel Region-Based Bayesian Approach for Genetic Association with Next Generation Sequencing (NGS) Data

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Background: NGS

Next Generation Sequencing Studies

- The emergence of new **high-throughput genotyping technologies**, such as Next Generation Sequencing (NGS), allows the study of the human genome at an unprecedented depth and scale
- They provide invaluable opportunities to decipher the **biological processes involved in complex human diseases**
- The study of the genetic landscape of **inherited and acquired mutations** in cancer patients could provide invaluable insights into the essential pathways driving the progression from a normal cell to non-invasive precursor lesions, and then to advanced and metastatic diseases

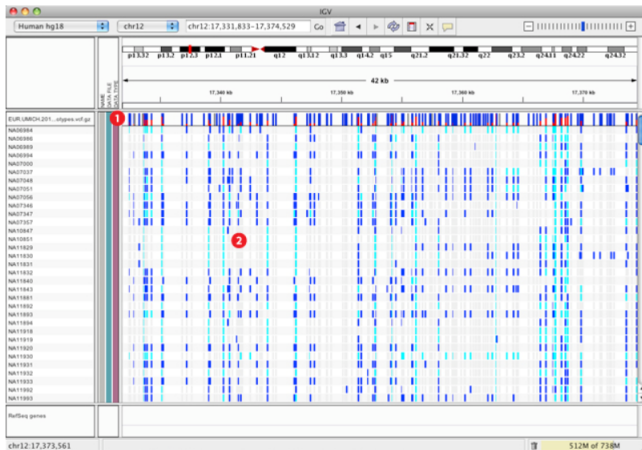
Outline of our framework

- Model setting
- Bayes Factor derivation for case-control design
- Prior definition
- Hyper-parameter specification
- Asymptotic properties
- Genome-wide inference
- Simulations with the program sim1000G
- Application on lung cancer study

Example NGS data

NGS data

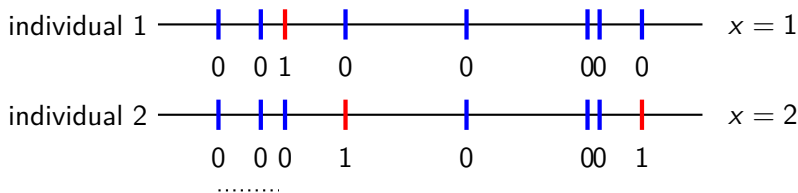
An example of sequenced genomic region is displayed below through the sequence viewer IGV.



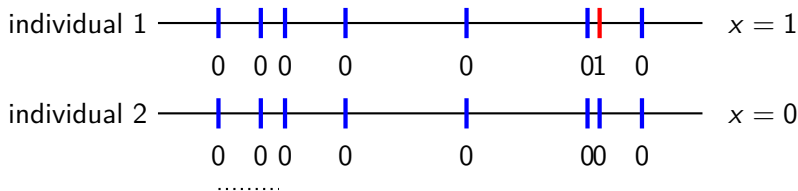
- 1 Each bar across the top of the plot shows the allele fraction for a single locus.
- 2 The genotypes for each locus in each sample. Dark blue = heterozygous, Cyan = homozygous variant, Grey = reference. Filtered entries are transparent.

Data example: a genetic region with 10 loci

Cases:



Controls:



Blue: non-mutated locus

Red: mutated locus

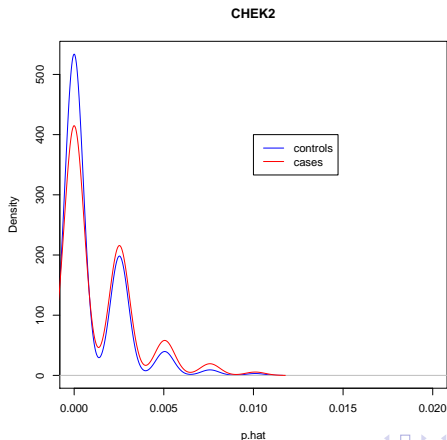
Density curve of \hat{p} of real data

k : individual k

n : number of loci in the region

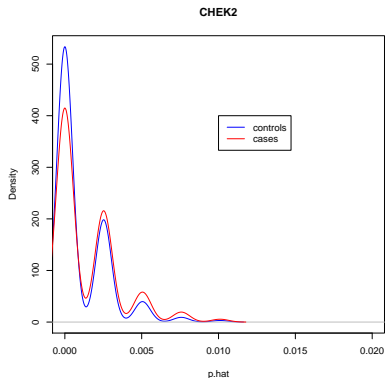
x_k : number of rare variants in the region for individual k , $x_k \sim \text{Binomial}(n, p_k)$

p_k : probability of having a rare variant at single locus for individual k , $\hat{p}_k = \frac{x_k}{n}$



Rationale for our rare variant association test

- **Goal:** Develop regional association test based on the comparison of rare variant rate (p_i) distribution between cases and controls.
- This comparison is accomplished by using the **Bayes Factor** (BF) statistic.

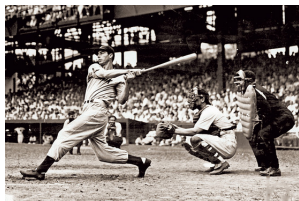


Background: Bayes Factor

Influential work on the BF: The "BayesBall"

- **Albert, J.** (2008), "Streaky Hitting in Baseball", Journal of Quantitative Analysis of Sports, vol. 4.
- **Albert, J.** (2013), "Looking at Spacings to Assess Streakiness", Journal of Quantitative Analysis of Sports, vol. 9.

Joe DiMaggio



- Bayes factor in support of true streakiness is

$$BF_{\kappa} = \frac{f(y|M_{\kappa})}{f(y|M)}.$$

BF in genetic association studies

- First **GWAS** application = the WTCCC study (2007)
- Some **review** in Stephens and Balding (Nat. Rev. Genetics, 2009)
- Wakefield (2009) formalized the BF in the context of GWAS
 - Interesting discussion about **informative priors** (effect-MAF dependence) vs. **non-informative** priors (implicit p-value prior)
 - Sketches the use of BF in the Bayesian False Discovery although not detailed
- **McCallum and Ionita-Laza, Biometrics 2015**

Methods

Model Setting

- Let X_{ijk} be the count of rare variants in the region i , for group j and individual k

$$X_{ijk} \sim \text{Binomial}(n_{ijk}, p_{ijk})$$

- Suppose that p_{ijk} varies across genetic regions and individuals, according to a prior density function $g(p_{ijk}|\theta_{ij})$, with $\theta_{ij} \equiv \theta_{i1}$ if j is in the control group and $\theta_{ij} \equiv \theta_{i2}$ if j is in the case group.
- Our goal is to assess whether there is a difference in rare variant counts between cases and controls in a particular region i by comparing : $H_{i0} : \theta_{i1} = \theta_{i2} = \theta_i$ vs. $H_{i1} : \theta_{i1} \neq \theta_{i2}$ using the Bayes Factor (BF) statistic.

- **Bayes Factor** (BF) is the ratio between the probabilities of the data (**marginal likelihood**) under the **alternative** hypothesis (association exists) and the **null** hypothesis (no association).

$$BF = \frac{m_1(X)}{m_0(X)}$$

- The marginal likelihood function under H_0 and H_1 :

$$m_0(X) = \int_P f(X|P)g(P)dP = \int_P f(X|P) \int_{\theta} g(P|\theta)\pi(\theta|\eta^*, K^*)d\theta dP$$

$$m_1(X) = \int_{P_1} f(X_1|P_1) \int_{\theta_1} g(P_1|\theta_1)\pi(\theta_1|\eta_1^*, K_1^*)d\theta_1 dP_1 \times \int_{P_2} f(X_2|P_2) \int_{\theta_2} g(P_2|\theta_2)\pi(\theta_2|\eta_2^*, K_2^*)d\theta_2 dP_2$$

where θ is the parameter we want to compare between cases and controls.

- There are **two definitions** for the prior distribution $g(P|\theta)$.

Prior definition I

- Under the **beta prior distribution**, we have

$$p_{ijk} | \theta_{ij} \sim \text{Beta}(\eta_{ij}, K_i),$$

Here the beta distribution is parametrized in terms of **mean** (denoted by η_{ij}) and **precision** (denoted by K_i). Relationship with (α, β) :

$$\eta = \frac{\alpha}{(\alpha + \beta)}, \quad K = \alpha + \beta.$$

- With the Beta prior, the **marginal distribution** of rare variants count in the region is **Beta-Binomial (BB)**. It assumes a similar pairwise correlation between the rare variants within the region. Our simulation studies (thanks to **Fode Tounkara**) showed that the BB fits the sequencing rare variants data much better than many Copula alternatives.

Prior definition II

- Under the **mixture prior distribution**, we assume that p_{ijk} follows a mixture distribution of a point mass at zero and a beta distribution with probability w_{0ij} and $w_{1ij} = 1 - w_{0ij}$, respectively:

$$X_{ijk} = \begin{cases} 0, & \text{if } p_{ijk} = 0 \text{ with } P(p_{ijk} = 0) = w_{0ij} \\ X_{ijk} \sim \text{Bin}(n_{ijk}, p_{ijk}), & \text{if } p_{ijk} > 0 \text{ with } P(p_{ijk} > 0) = 1 - w_{0ij} \end{cases}$$

Also when $p_{ijk} > 0$, the prior density for p_{ijk} is **Beta**(η_{ij}, K_i).

Hierarchical hyper-parameter specification

- Our **hyper parameters of interest** are η , η_1 , η_2 , w_{01} , w_{02} , and w_0 .
- We assume a **hierarchical prior structure** where each hyper-parameter is assumed to follow a **beta distribution** with new mean and precision parameters η^* , η_1^* , η_2^* , K^* , K_1^* , K_2^* .
- The parameters of the prior and hyperprior distributions are estimated empirically from the data by using **MLE**.

BF distribution under the null

- Ideal parameters η^* and K^* should lead to:
 - BF is **independent of gene size**
 - BF ($\log BF$) has a **known theoretical distribution**
- *Theorem 1.* Assume that $\eta^* = \hat{\eta}$, $K^* = \hat{\eta} \hat{\Sigma}^{-1}$, $\eta_1^* = \hat{\eta}_1$, $K_1^* = \hat{\eta}_1 \hat{\Sigma}_1^{-1}$, $\eta_2^* = \hat{\eta}_2$ and $K_2^* = \hat{\eta}_2 \hat{\Sigma}_2^{-1}$, for gene i , when sample size $N_1 \rightarrow \infty$ and $N_2 \rightarrow \infty$,

$$2 \log BF = \frac{(\hat{\eta}_1 - \hat{\eta}_2)^2}{\hat{\Sigma}_1 + \hat{\Sigma}_2} \sim \chi^2(1)$$

BF with individual-level covariates

For group j ($j=1$ or 2 , $j=1$, control group, $j=2$, case group), individual k , $p_{jk} \sim \text{Beta}(\eta_{jk}, K)$. We build Beta regression to model the relationship between covariate vector w_{jk} with length equal to c and the rare variant rate at single locus p_{jk} .

- Version 1

$$\text{logit}(\eta_{jk}) = \beta_{0j} + w_{jk}\beta$$

$$\beta_{0j} \sim \text{Normal}(\mu_j, \sigma_j^2)$$

$$\beta \sim \text{MVN}(\mu_\beta, B)$$

- Version 2

$$\text{logit}(\eta_{jk}) = \text{logit}(\eta_j) + R_{jk},$$

where $R_{jk} = \beta w_{jk}$ and w_{jk} is a vector of PCs or ethnic group indicator variables.

$$\eta_j \sim \text{beta}(\eta_j^*, K_j^*)$$

Bayesian FDR

Bayesian control of False Discovery Rate (FDR) for genome wide inference

- The goal of **genome-wide inference** is to perform a simultaneous testing of multiple hypotheses (i.e. all the genes or genomic regions) = **m null hypotheses** $H_i, i = 1, \dots, m$, using data Y
- Let $Z_i = 1$ if **H_i is true** and $Z_i = 0$ if **H_i is false**, $i = 1, \dots, m$, and π_0 the proportion of regions/genes generated under the null
- We have $Z_i | \pi_0 \sim \text{Bernoulli}(1 - \pi_0)$
- We also define δ_i denote a **decision rule** in $(0, 1)$ on Z_i based on the data and $D = \sum_{i=1}^m \delta_i$

Bayesian control of False Discovery Rate (FDR) for genome wide inference

Following Muller et al. (2006), the **False Discovery Proportion (FDP)** is defined as

$$\text{FDP} \equiv \frac{\sum_{i=1}^m \delta_i (1 - Z_i)}{D \vee 1},$$

and the **Bayesian FDR** as:

$$\overline{\text{FDR}} \equiv E(\text{FDP} | Y) = \frac{\sum_{i=1}^m \delta_i (1 - v_i)}{D \vee 1}.$$

The interest in the **Bayesian control of the FDR**, is to estimate $v_i \equiv \text{Pr}(Z_i = 1 | Y)$ by

$$\hat{v}_i = \frac{(1 - \hat{\pi}_0) BF_i}{\hat{\pi}_0 + (1 - \hat{\pi}_0) BF_i}$$

Estimate of $\hat{\pi}_0$

- Wen et al. (2016) showed that an **upper bound** estimation of π_0 can be obtained by

$$\hat{\pi}_0 = \frac{\sum_{i=1}^m I(BF_i \leq q_{i,\gamma})}{m\gamma}.$$

=> requires permutations to assess the null distribution of the BF for each gene

=> lacks well study of impact of γ

- Since we proved that $2 \log BF_i \xrightarrow{d} \chi^2(1)$, we can then **estimate** π_0 by

$$\hat{\pi}_0 = \frac{\sum_{i=1}^m I(2 \log BF_i \leq q_\gamma^*)}{m\gamma},$$

where q_γ^* is the γ -quantile of a $\chi^2(1)$ distribution

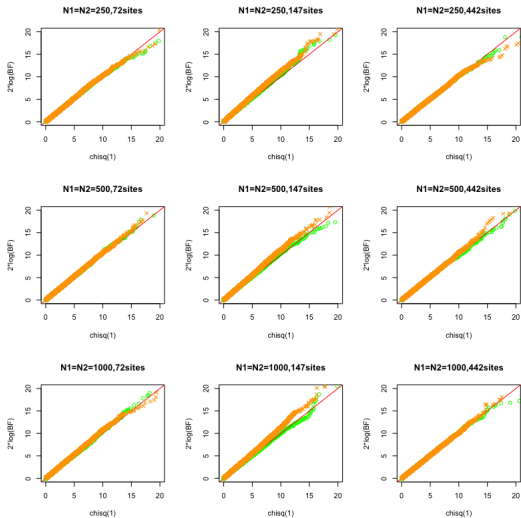
=> which avoids the need for permutations

=> Try to find optimal value of γ

Simulation Procedure

- R package "**sim1000G**" is used to simulate the rare variant genotype data.
 - Now available on the CRAN, credit to **Apostolos Dimitromanolakis**
 - The simulated data can capture the **allele frequencies** and **LD patterns** in the genome, as well as **recombination hotspots**.
 - Only choose variants with $MAF \in (1e - 6, 0.01)$ for data analysis.
- Number of **causal variants** is proportional to the region size. We assume all causal variants are **deleterious**, with **OR** = 2.63 to 3.73, inversely related to MAF.
- Each simulated dataset has same number of cases and controls.

QQ plot: BF simulated under the null



Simulation Results

Table: Statistical power of different methods for different gene sizes and sample sizes with 1,000 replicates (reject null hypothesis when $p < 0.05$)

Statistical Test		$N_1 = N_2 = 250$			$N_1 = N_2 = 500$			$N_1 = N_2 = 1000$		
		72 sites	147 sites	442 sites	72 sites	147 sites	442 sites	72 sites	147 sites	442 sites
BF method										
Beta prior	Compare η	23.8	41.3	87.2	35.3	58.9	98.3	59.4	82.2	100.0
Mixture prior	Compare η	25.6	44.4	88.7	37.1	61.7	98.2	62.2	83.5	100.0
SKAT		13.1	22.0	50.2	24.9	45.2	86.1	55.7	79.1	99.9
Burden		16.9	30.2	83.5	25.8	50.2	96.6	48.4	75.1	100.0
SKAT-O		16.8	32.6	82.5	29.9	57.6	98.0	61.8	88.5	100.0

Lung cancer data application

Lung Cancer Study

- Our data is from lung cancer exome-sequencing consortium study, including 4 different cohorts.
- After removing the duplicated individuals, sample size of different cohorts

Cohort	cases	controls	Total
Toronto	260	258	518
Liverpool	65	69	134
HSPH	426	269	695
IARC	293	284	577

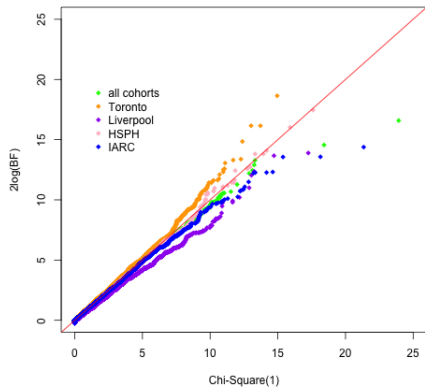
- After filtering out multi-allelic variants, the **MAF distribution** for the bi-allelic variants are

MAF	0	(0,0.01]	(0.01,0.05]	(0.05,0.5]	Total
#(Variants)	62,940	1,095,794	60,204	129,412	1,348,350
Proportion (%)	4.7	81.3	4.5	9.6	

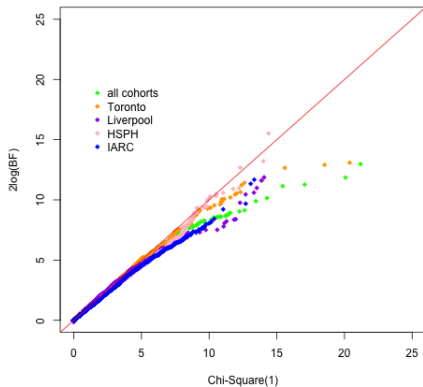
- In the analysis, the number of sites within the gene is at least **20** for **beta prior BF** and **50** for **mixture prior BF**.
- The **number of genes** used for beta prior BF and mixture prior BF are 14,321 and 7,454 respectively.

QQ plot: include all variants

QQ plot for beta prior BF

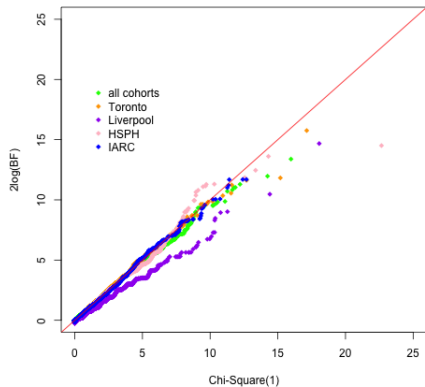


QQ plot for mixture prior BF

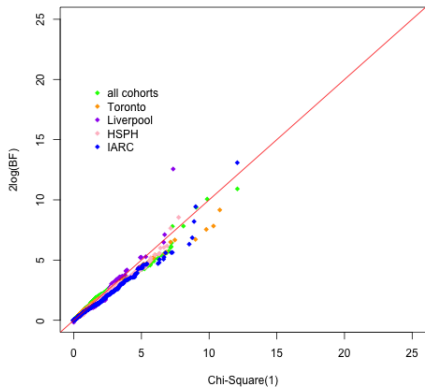


QQ plot: include high impact variants

QQ plot for beta prior BF

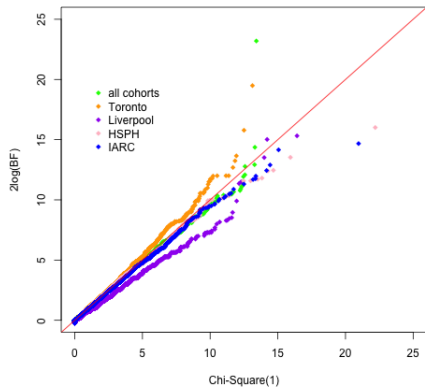


QQ plot for mixture prior BF

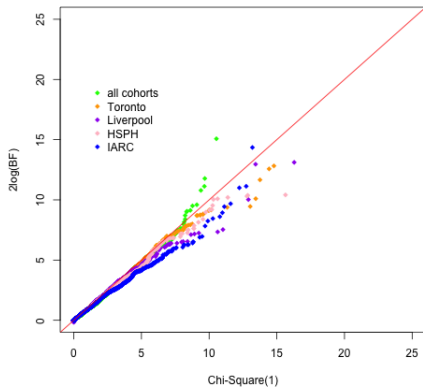


QQ plot: include high and moderate impact variants

QQ plot for beta prior BF



QQ plot for mixture prior BF



Bayesian FDR application

Table: Estimate of π_0

	$\gamma = 1 - \frac{1}{m}$		$\gamma = 0.99$		$\gamma = 0.95$		$\gamma = 0.9$	
	beta	mixture	beta	mixture	beta	mixture	beta	mixture
all variants	1	1	0.9993095	1	1	1	1	1
high risk	1	1	0.9995661	1	1	1	0.9952272	1
moderate risk	1	1	1	1	0.9987782	1	0.9992063	1

FDR of the top gene using beta prior in the moderate risk dataset:

- $\gamma = 0.95$, FDR ≈ 0.007
- $\gamma = 0.9$, FDR ≈ 0.01

Top 20 genes with beta prior: high impact variants

gene.name	chr	sites	BFbeta	p.beta	BF(TO)	BF(Livepool)	BF(HSPH)	BF(IARC)
CAMTA2	17	48	807.97	2.53e-04	35.13	1.64	1.71	10.48
ADAMTSL4	1	52	397.20	5.41e-04	3.85	0.98	17.91	10.09
CACNA1G	17	44	283.65	7.77e-04	2.54	0.97	13.08	10.24
SCRIB	8	56	249.19	8.93e-04	4.35	1.70	2.19	24.98
SREBF2	22	43	247.25	9.01e-04	8.26	0.94	3.38	21.21
ERBB2	17	36	224.39	1.00e-03	7.70	0.89	5.06	2.10
PCDH7	4	22	212.20	1.06e-03	4.62		5.35	3.61
SAMD4B	19	21	139.04	1.68e-03	1.16			1.95
CDC42BPA	1	38	135.67	1.73e-03	1.03		1.15	346.24
PAMR1	11	22	127.98	1.84e-03	7.83		1.05	24.03
PP2D1	3	31	121.32	1.95e-03	2.45	2.21	11.00	3.03
WDR92	2	21	120.58	1.96e-03		1.65	1.08	7.29
CCDC60	12	31	116.36	2.04e-03	9.69	0.89	1414.61	1.04
ABL2	1	30	114.33	2.08e-03	5.43	3.52	1.66	4.98
KIF20A	5	24	113.20	2.10e-03	49.67		2.74	2.92
RBM14	11	21	110.41	2.16e-03		1.09	5.84	1.77
TERT	5	26	106.05	2.26e-03	28.90	0.89	1.39	13.90
AXDND1	1	37	90.50	2.68e-03	1.30	0.94	11.12	5.40
LRSAM1	9	37	86.92	2.81e-03	195.87	2.64	1.91	1.41
FN1	2	63	78.13	3.15e-03	2.51	1.07	5.26	3.70

Impact of protective variants

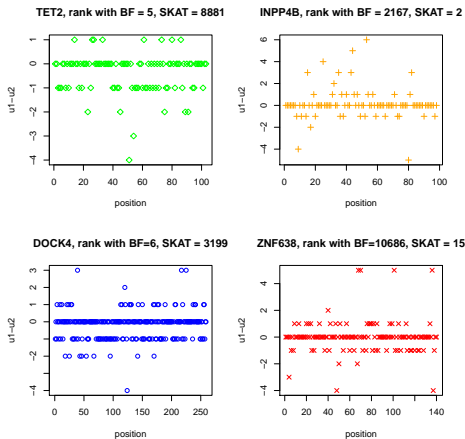


Figure: The Y-axis represents the difference in total minor allele counts between controls (u_1) and cases (u_2) at each single site (locus) of the region. If the genetic variant has a deleterious effect on the disease, then $u_2 > u_1$ and conversely if it has a protective effect, then $u_1 > u_2$.

Discussion

- The use of **empirical Bayes priors along with a Bayesian control of FDR** offer a comprehensive framework to make genome-wide statistical inference about the important chromosomal regions associated with the disease of interest
- **How to define the priors?** asymptotic properties of BF or informative priors?
- $\log\text{BF} \approx \log\text{LR} + \log \frac{\pi(\theta|H_1)}{\pi(\theta|H_0)}$ - term
- The **regression framework** might offer a good compromise (Zhou and Guan, JASA, 2018) but still not fully developed for discrete outcomes
- **Future developments** include the extension of the BF approach to account for variant-level covariates and family designs

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